



September 1, 2026

Mr. William Nelson
Safety Coordinator
Pender County Schools
925 Penderlea Highway
Burgaw, North Carolina 28425

Re: INDOOR AIR QUALITY SCREENING
Pender County Schools – South Topsail Elementary School
17385 US-17
Hampstead, North Carolina 28443
EI Project No. IHMO260169.00

Mr. Nelson:

The EI Group, Inc. (EI) is pleased to present this report for the Indoor Air Quality (IAQ) Screening performed for Pender County Schools at South Topsail Elementary School located at 17385 US-17 in Hampstead, North Carolina. EI was requested to perform a limited IAQ assessment to include fungal air sampling and visual assessment within Room 108 (classroom) of the school.

Project Information

Mr. Thomas Majkowski, Industrial Hygienist with The EI Group, Inc., performed the site assessment on August 28, 2026. The purpose of the assessment was to perform an IAQ screening within the assessed area of the school building to include a limited visual and moisture survey.

Site Observations

The weather on this day was sunny, hot and humid. Mr. Majkowski visually assessed Room 108 inside the school for visible/accessible signs of elevated moisture conditions and/or water damage, such as stains, discoloration, rust on metal, musty odors, and suspect visible mold growth. EI also looked for conditions that may lead to mold growth, such as high humidity, standing water, and wet or damp building materials.

Air scrubbers and dehumidifiers were noted within the classroom. The machines were not operating during the assessment. Portions of the contents within the classroom were stored in plastic bags.

Moisture Survey/ Visual Observations:

Investigation was conducted by utilizing an infrared (IR) camera and a moisture meter to identify building materials wet enough to be “at risk” for fungal growth.

No suspect visible fungal growth or elevated moisture conditions were observed within accessible areas of the classroom at the time of assessment. The Diffusers and Return within the classroom were clean with no visible dust or suspect microbial growth.

Sampling

Direct Reading Instruments

Temperature and relative humidity (RH) readings were collected using a Fluke 975 Air Meter to determine if the air-handling systems were operating effectively to condition the assessed spaces for occupant comfort within the interior of the building. These parameters are also analyzed to determine if conditions are being maintained to minimize the potential for fungal growth and amplification in the office environment. Measurements are compared to a range, recommended by the American Society of Heating, Refrigeration, and Air-conditioning Engineers (ASHRAE). ASHRAE Standard 55, “Thermal Environmental Conditions for Human Occupancy”, generally recommend maintaining a temperature range anywhere between 67°F and 82°F and maintaining a relative humidity range below 60%. Specific temperature comfort levels are dependent on relative humidity, season, clothing worn, and other factors. During the summer months thermal comfort is usually achieved at the upper end of the listed range.

Results from monitoring indicated indoor temperature was within standard comfort conditions within Room 108, measured at 77.3°F. The outdoor temperature ranged averaged 79.1°F during the assessment.

RH readings were within recommended guidelines in Room 108, measured at 45.7%. The outdoor RH levels averaged 90.0% during the assessment.

Air Sampling Methods and Results

Spore Trap Air Sampling

Sampling for airborne fungal spores was conducted within Room 108 to determine airborne fungal counts, the genera present, and the relative levels of each genus. Two (2) air samples were also collected from outdoors as reference samples.

Air sampling was conducted utilizing Air-O-Cell® “spore-trap” sampling media. All air samples were collected with a Buck Pump™ sampling pump, calibrated with a bubble meter which was calibrated from a primary standard. The Buck sampling pump was calibrated to 15 liters per minute (LPM) with a sampling time of 5 minutes, giving a total sample volume of 75 liters. Following the collection of fungal samples, sampling media were numbered consecutively, placed under chain-of-custody, and transported to EMSL, in Morrisville North Carolina for analysis by direct read microscopic screening. Spore identification of fungal air samples was performed and reported by the analytical laboratory. The results of analysis, along with a copy of the sample chain-of-custody, are in appendices at the conclusion of this report.

Fungal air sampling results indicated that the airborne fungal concentrations collected in this space were lower in total and individual spore concentrations, in comparison to the outdoor reference levels. In

addition, spore genera collected in the indoor samples were similar to those in the outdoor reference samples.

Conclusions and Recommendations

No suspect visible fungal growth or elevated moisture conditions were observed within accessible areas of the classroom at the time of assessment. The Diffusers and Return within the classroom were clean with no visible dust or suspect microbial growth.

Temperature readings were within recommended guidelines within Room 108 at the time of assessment.

Fungal air sampling results indicated that the airborne fungal concentrations collected in this space were lower in total and individual spore concentrations, in comparison to the outdoor reference levels. In addition, spore genera collected in the indoor samples were similar to those in the outdoor reference samples.

Best Practices:

1. Change HVAC filters per manufacturers recommended intervals and general housekeeping on a set schedule limiting airborne dust loading on horizontal surfaces.
2. A comprehensive mold/moisture assessment of Room 108 should be performed to include ceiling plenums, plumbing penetrations (bathroom), HVAC Systems, and the exteriors.

Qualifications and Limitations

This report summarizes our evaluation of the conditions observed at the above-mentioned site. Our findings are based upon our observations of the assessed areas within South Topsail Elementary School located in Hampstead, North Carolina, and results of analyses of the samples obtained at the time of our visit only. Any conditions discovered which deviate from the data contained in this report should be presented to EI for our evaluation. The IAQ screening was limited to the observed areas within the classrooms and did not include the bathroom/plumbing penetrations, ceiling plenums, and the exteriors.

It should be noted that successful remediation of visible growth does not guarantee that the mold will not return or that all indoor sources of mold have been discovered or removed. Molds are part of the natural environment and can grow on virtually any surface if adequate moisture, nutrients, and oxygen are present. It is therefore impossible to eliminate all molds and mold spores indoors. Mold amplification, however, can be controlled by controlling moisture indoors. If remediation of visible fungal growth is performed, the goal of mold remediation is to remove or clean the contaminated materials in a way that prevents emission of mold and dust contaminated with mold from leaving the work area and entering an occupied or non-remediation area, while protecting the health of workers performing remediation. The work is typically accomplished by trained workers using polyethylene barriers, exhaust fans equipped with HEPA filters, HEPA vacuums, damp wiping, and personal protective equipment. To fully accomplish

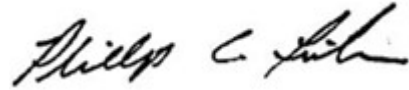
the fungal remediation process, the underlying source(s) of water accumulation or moisture must be rectified, or mold growth will recur.

This report is intended for the exclusive use of Pender County Schools. Other parties shall not rely upon this report and the findings herein without prior written consent of The EI Group, Inc. The findings are relative to the date of our site visit and should not be relied upon for substantially later dates.

Sincerely,
The EI Group, Inc.



Thomas Majkowski
Manager, Industrial Hygiene Services



Phil Fincher, CIH, CSP
Senior Vice President

Attachments:

- Laboratory Analysis Results

Laboratory Analysis Results

Spore Trap Sampling



EMSL Analytical, Inc.

2500 Gateway Centre Blvd., Suite 600 Morrisville, NC 27560

Tel/Fax: (919) 465-3900 / (919) 465-3950

<http://www.EMSL.com> / raleighlab@emsl.com

EMSL Order: 292604682

Customer ID: ENVI67

Customer PO:

Project ID:

Attention: Thomas Majkowski

The EI Group, Inc.

2101 Gateway Centre Blvd, Suite 200

Morrisville, NC 27560

Phone: (919) 657-7500

Fax: (919) 657-7551

Collected Date: 08/28/2026

Received Date: 08/31/2026 09:50 AM

Analyzed Date: 08/31/2026

Project: IHMO260169.00

Test Report: Air-O-Cell(™) Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)

Lab Sample Number:	292604682-0001			292604682-0002			292604682-0003		
Client Sample ID:	A1			A2			A3		
Volume (L):	75			75			75		
Sample Location:	Outdoor 1			Room 108			Outdoor 2		
Spore Types	Raw Count†	Count/m³	% of Total	Raw Count†	Count/m³	% of Total	Raw Count†	Count/m³	% of Total
Alternaria (Ulocladium)	1	40	0.1	1	10*	2.8	-	-	-
Ascospores	56	2400	3.1	-	-	-	60	2500	3.2
Aspergillus/Penicillium++	1	40	0.1	-	-	-	14	590	0.8
Basidiospores	108(1760)	74300	95	8	300	83.3	107(1740)	73400	95.4
Bipolaris++	-	-	-	-	-	-	-	-	-
Chaetomium++	-	-	-	-	-	-	-	-	-
Cladosporium	28	1200	1.5	1	40	11.1	8	300	0.4
Curvularia	-	-	-	1	10*	2.8	2	30*	0
Epicoccum	-	-	-	-	-	-	-	-	-
Fusarium++	-	-	-	-	-	-	-	-	-
Ganoderma	4	200	0.3	-	-	-	3	100	0.1
Myxomycetes++	2	30*	0	-	-	-	-	-	-
Pithomyces++	-	-	-	-	-	-	1	40	0.1
Rust	-	-	-	-	-	-	-	-	-
Scopulariopsis/Microascus	-	-	-	-	-	-	-	-	-
Stachybotrys/Memnoniella	-	-	-	-	-	-	-	-	-
Unidentifiable Spores	-	-	-	-	-	-	-	-	-
Zygomycetes	-	-	-	-	-	-	-	-	-
Total Fungi	1852	78210	100	11	360	100	1828	76960	100
Hyphal Fragment	-	-	-	1	40	-	-	-	-
Insect Fragment	-	-	-	-	-	-	-	-	-
Pollen	-	-	-	-	-	-	-	-	-
Analyt. Sensitivity 600x	-	42	-	-	42	-	-	42	-
Analyt. Sensitivity 300x	-	13*	-	-	13*	-	-	13*	-
Skin Fragments (1-4)	-	1	-	-	3	-	-	1	-
Fibrous Particulate (1-4)	-	-	-	-	1	-	-	1	-
Background (1-5)	-	1	-	-	2	-	-	1	-

† Due to method stopping rules, extrapolated raw counts are reported in parenthesis.

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

Billy Barnes, Laboratory Manager
or other Approved Signatory

No discernable field blank was submitted with this group of samples.

EMSL Analytical, Inc. maintains liability limited to cost of analysis. Interpretation and use of test results are the responsibility of the client. This report relates only to the samples reported above, and may not be reproduced, except in full, without written approval by EMSL Analytical, Inc. EMSL Analytical, Inc. bears no responsibility for sample collection activities or analytical method limitations. The report reflects the samples as received. Results are generated from the field sampling data (sampling volumes and areas, locations, etc.) provided by the client on the Chain of Custody. Samples are within quality control criteria and met method specifications unless otherwise noted. Skin Fragment and Fibrous Particulate ratings are based on the percent of non-fungal material they represent: 1 (1-25%), 2 (26-50%), 3 (51-75%), or 4 (76-100%). Background ratings are based on the total area covered by non-fungal particles: 1 (1-25%), 2 (26-50%), 3 (51-75%), 4 (76-99%), or 5 (100%; overloaded). High levels of background particulate can obscure spores and other particulates, leading to underestimation. Background levels of 5 indicate an overloading of background particulates, prohibiting accurate detection and quantification. Present = Spores detected on overloaded samples. Results are not blank corrected unless otherwise noted. The detection limit is equal to one fungal spore, structure, pollen, fiber particle or insect fragment. "*" Denotes particles found at 300X. "-" Denotes not detected. Due to method stopping rules, raw counts >= 100 are extrapolated based on the percentage analyzed.

Samples analyzed by EMSL Analytical, Inc. Morrisville, NC AIHA LAP, LLC-EMLAP Accredited #173741

Initial report from: 09/01/2026 07:09 AM

For information on the fungi listed in this report, please visit the Resources section at www.emsl.com

Microbiology Chain of Custody Form

The EI Group, Inc.

IHMO260169.00

8/31/2026 9:50

M001 Air-O-Cell

TAT: 24 Hour
Air

Order ID: 292604682

No Samples: 3

Due: 09/01 9:50 AM

Fax: 919-657-7551

:7560

(919) 465-3900

sighlab@emsl.com

written authorization.

EMSL
LABORA

Customer Information	Customer ID	Billing ID:
	Company Name: The EI Group, Inc.	Company Name: The EI Group, Inc.
	Contact Name: Thomas Majkowski	Billing Contact: Thomas Majkowski
	Street Address: 2101 Gateway Centre Blvd Suite 200	Street Address: 2101 Gateway Centre Blvd, Suite 200
	City, State, Zip Code: Morrisville NC 27560 Country: US	City, State, Zip Code: Morrisville NC 27560 Country: US
	Phone: 9199064686	Phone: 9199064686
Email(s) for Report: tmajkowski@ei1.com	Email(s) for Invoice: akuttles@ei1.com; tmajkowski@ei1.com	

Project Information

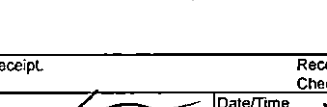
Project Name/No: IHMO260169.00	Purchase Order:
EMSL LIMS Project ID: (If applicable, EMSL will provide)	State of Connecticut (CT) must select project location: ☐ Commercial (Taxable) ☐ Residential (Non-taxable)
State Samples Collected: NC	Zip Code Samples Collected:
Sampled By Name:	Sampled By Signature:
	Sampler ID# (If applicable)
	No. of Samples in Shipment: 3
Sterile, Sodium Thiosulfate Preserved Bottle Used: ☐ Biocide Used In Source (specify)	
Public Water Supply Samples: ☐ Note: All results may automatically be reported to DOH if required by State.	
Turn-Around-Time (TAT) Please call ahead for large projects and/or turn-around times 6 Hours or Less. *32 Hour TAT available for select tests only; samples must be submitted by 11:30am.	
☐ 3 Hour ☐ 6 Hour ☒ 24 Hour ☐ 32 Hour ☐ 48 Hour ☐ 72 Hour ☐ 96 Hour ☐ 1 Week ☐ 2 Week	

MICROBIOLOGY TEST CODES

M001 Air-O-Cell	M174 MoldSnap	M009 Bacteria Culture Gram Stain & Count	M115 Sewage Screen - Water (P/A***)
M030 MICRO 5	M032 Allergenco-D	M010 Bacteria Count & ID - 3 Most Prominent	M116 Sewage Screen - Water (MPN**)
M041 Fungal Direct Examination		M011 Bacteria Count & ID - 5 Most Prominent	M117 Sewage Screen - Swab (P/A***)
M169 Pollen ID & Enumeration			M013 Sewage Screen - Swab (MFT*)
M005 Viable Fungi-Air Samples (Genus ID & Count)		M012 Pseudomonas aeruginosa (P/A***)	M730 Methicillin-resistant Staph. aureus (MRSA)
M006 Viable Fungi-Air Samples (Includes Penicillium, Aspergillus, Cladosporium, Stachybotrys Species ID & Count)		M024 Pseudomonas aeruginosa (MFT*)	M031 Rapid-growing non-TB Mycobacteria Detection & Enumeration
M007 Culturable Fungi-Surface Samples (Genus ID & Count)		M015 Heterotrophic Plate Count	M014 Endotoxin Analysis
M008 Culturable Fungi-Surface Samples (Includes Penicillium, Aspergillus, Cladosporium, Stachybotrys Species ID & Count)		M017 Total Coliform & E. Coli (Colilert P/A***)	M044 Group Allergen (Cat, Dog, Cockroach, Dust Mite)
M280 Dust Characterization Level-1		M018 Total Coliform & E. Coli (MFT*)	M095 Bacteroides
M281 Dust Characterization Level-2		M114 Total Coliform & E. Coli Enumeration (Colilert MPN**)	Other - See Analytical Price Guide for Test Code
Add On to Spore Trap & M041 Analyses		M019 Fecal Coliform (MFT*)	Legionella Analysis Please use EMSL Legionella COC
available at certain lab locations		M020 Fecal Streptococcus (MFT)	
M280A Dust Characterization Level-1		M029 Enterococci (MFT*)	
M281A Dust Characterization Level-2		M129 Enterococci (Enterolert P/A***)	*MFT= Membrane Filtration Technique
		M180 Real Time qPCR-ERMI 36 Panel	**MPN = Most Probable Number
		M025 Sewage Screen - Water (MFT*)	***P/A = Presence/Absence

Sample #	Sample Location/Description	Sample Type (Matrix)	Potable / Non-Potable (Only for Water)	Test Code	Volume/Area	Date / Time Collected	Temperature (Lab Use Only)
A1	Outdoor 1	Air		M001	75L	8-28-26	
A2	Room 108	Air		M001	75L	8-28-26	
A3	Outdoor 2	Air		M001	75L	8-28-26	

Special Instructions and/or Regulatory Requirements (Sample Specifications, Processing Methods, Limits of Detection, etc.)

Method of Shipment:	Sample Condition Upon Receipt:	Received on Ice? Check if Yes: ☐
Relinquished by: 	Date/Time: 8-28-26	Received by: 
Relinquished by:	Date/Time:	Received by:
		Date/Time: 8/31/26 9:50

Controlled Document - COC-34 Micro R18 02/13/2026

☐ AGREE TO ELECTRONIC SIGNATURE (By checking, I consent to signing this Chain of Custody document by electronic signature.)

EMSL Analytical, Inc.'s Laboratory Terms and Conditions are incorporated into this Chain of Custody by reference in their entirety. Submission of samples to EMSL Analytical, Inc. constitutes acceptance and acknowledgment of all terms and conditions by Customer.