PINNACLE CONSULTANTS

10 Independent Avenue
Nitro, West Virginia 25143

Office: 304-757-5204
Fax: 304-440-3465

www.pinnaclecorp.net

INDOOR AIR QUALITY ASSESSMENT

AT

Ceredo-Kenova Elementary
300 9th Street
Kenova, WV 25530

Prepared for

Custom Services Industries
Mr. Michael McCallister
533R 3rd Avenue
Huntington, WV 25701

September 16, 2026

Project ID: 26-654

Helping you understand your Environment because...

What you don't know ***CAN*** hurt you!

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Pinnacle Consultants (Pinnacle) conducted a mold assessment at the above reference address. The purpose of this inspection was to determine if fungal levels were elevated at the time of sample collection.

For this project, a total of sixteen (16) indoor air samples and one (1) tape-lift sample were collected, plus an additional exterior sample for comparison. Air samples were collected at the following locations: Rooms 238, 231 (Janitor Closet), 224, 217, 212, Hall @ 190, 182, 144, 180, 169, 159, 178 (Library Containment), Cafeteria/124, 111, 142, and the Clinic, along with an exterior sample for comparison. A tape-lift sample was collected from the supply vent in the Clinic to confirm the presence of biological material and determine the level of fungal spores.

Findings

Observations made at the time of testing did not identify any active water intrusion, standing moisture, or visible microbial growth in the areas observed at the time of testing. Relative humidity readings collected throughout the building ranged from 46.1% to 58.9%, within the normal range for a properly functioning HVAC system, and are not indicative of an elevated indoor moisture condition. Sample 654-2 (Room 231) was collected while the adjacent janitor's closet was being swept, and Sample 654-12 (Room 178) was collected within an active containment area set up in the Library; neither condition is believed to have measurably affected the sample results.

Indoor carbon dioxide (CO2) readings were meaningfully elevated above the exterior baseline (466 ppm) in four occupied classrooms – Room 182, Room 180, Room 169, and Room 159 – ranging from 1,566 to 1,755 ppm. Elevated CO2 indicates reduced fresh-air exchange relative to occupancy load and is a ventilation consideration independent of the fungal findings discussed below.

The laboratory analysis result for the air sampling indicated **no elevated** levels at the time of testing.

The laboratory analysis results for the tape-lift samples collected are below:

Sample # 654-18 Clinic Vent Tape - confirmed the presence of ***Cladosporium*** spores at a Moderate spore estimate with a Few (possible active growth) mycelial/hyphal estimate. This is one of the most common fungal genera found and is typically an allergen. *See Fungal Descriptions for full description.*

The presence of hyphae associated with these fungal structures indicates active fungal growth on the sampled surface rather than the simple deposition of airborne spores. Hyphae are the thread-like filamentous structures that make up the vegetative portion (mycelium) of a mold colony and are responsible for nutrient absorption and colony growth.

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It's important to note that individual sensitivity to mold spores can vary, and not everyone exposed to these molds will experience health effects. Those with pre-existing respiratory conditions, allergies, or compromised immune systems are generally more susceptible to mold-related health effects. If you suspect mold exposure and experience persistent health symptoms, consider consulting a medical professional for proper evaluation and guidance.

As part of the Indoor Air Quality investigation, relative humidity levels, carbon dioxide and temperatures are checked at the time of sample collection.

ASHRAE recommends a relative humidity (rH) between 20-60%. At the time of testing the humidity level **was within** the recommended range.

ASHRAE recommends the temperature be between 68°-79° Fahrenheit. At the time of testing the temperature **was within** the recommended range at nearly all locations, with two areas (Cafeteria/124 at 67.1°F and Room 111 at 67.5°F) reading nominally below range by less than one degree.

Carbon Dioxide (CO2) levels are utilized as an overall check on air exchanges and the ability of the system to bring fresh air into the area. ASHRAE recommends Carbon Dioxide levels do not exceed 700 above the outside levels. Carbon Dioxide level **exceeded** the recommended range in four occupied classrooms (Rooms 182, 180, 169, and 159), indicating reduced fresh-air ventilation relative to occupancy in those spaces; all other sampled locations were within the recommended range.

Location	Temp (F)	rH%	CO2
Room 238	71.6	52.2	814
Room 231 (Janitor Closet)	70.7	57.8	736
Room 224	70.9	55.2	737
Room 217	70.7	46.1	542
Room 212	71.2	48.5	648
Hall @ 190	71.1	51.0	844
Room 182	71.7	57.1	**1566**
Room 144	72.4	46.9	741
Room 180	70.9	57.8	**1478**
Room 169	70.8	56.0	**1606**
Room 159	72.3	58.3	**1755**
Room 178 (Library Containment)	76.8	52.6	990
Cafeteria / Room 124	67.1	50.1	624
Room 111	67.5	58.9	538
Room 142	73.5	47.5	965
Clinic	—	—	—
Outside (Exterior Comparison)	87.7	38.7	466

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RECOMMENDATIONS

The airborne mold spore concentrations collected throughout the building **did not indicate indoor fungal amplification when compared to the exterior sample**. The tape-lift sample collected from the Clinic supply vent confirmed the presence of Cladosporium at a Moderate spore estimate with a Few (possible active growth) mycelial estimate. This finding indicates a localized accumulation of fungal spores on the vent surface, most likely associated with settled dust and organic debris rather than active water intrusion – relative humidity readings collected throughout the building were within the normal range, and no evidence of active water intrusion was observed during the inspection. Given the limited scope and modest spore/hyphal estimates of this finding, Pinnacle's recommendation is focused on cleaning of the affected unit and louvers; broader remediation of adjacent building materials **is not** warranted by the current data.

Based upon the observations and analytical results Pinnacle makes the following recommendations. Pinnacle recommends all remediation be performed by properly trained and equipped professionals, following the **Institute of Inspection, Cleaning and Restoration Certification (IICRC) S520 Guidelines** and using HEPA-filtered air filtration devices (AFDs), air scrubbers and vacuums along with a proper containment system to prevent the mold spores from spreading to unaffected areas. Proper PPE including P100 equipped respirators should be used to protect the workers during cleaning. Prior to cleaning, any issues regarding excess moisture should be addressed to prevent further or additional contamination. In addition Pinnacle recommends:

a. No Federal, State or Local regulations exist regarding mold exposure levels or remediation requirements.

b. Any building material which may be disturbed during this project **must** be tested for asbestos content prior to disturbance, **regardless of construction or manufacture date** according to WV State regulation 64-CSR-63 and EPA regulations. The contractor must follow all EPA, OSHA, and State of WV Regulations regarding Lead Paint, Asbestos, Respiratory Protection, or any other applicable standard.

c. Pinnacle recommends the following products or similar for cleaning, and sealing of surfaces:

a. Benefect Botanical DECON 30 or ShockWave by Fiberlock Technologies for cleaning.
b. Bad Axe Particle Conqueror as a particle suppressant.
c. AfterShock or IAQ6000 by Fiberlock Technologies as a sealant.
d. A hydrogen peroxide or chlorine-based product may be used to help eliminate any staining. **Bleach is strongly discouraged.**

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e. Follow all manufacturers' instructions for any product being utilized for cleaning, particle suppressant or sealant.

d. Containment of the work area and airflow management is recommended but not required; the work area should be covered with plastic sheet(s) and sealed with tape before remediation, to contain dust/debris. All openings should be sealed, as required, with critical barriers constructed of 1 layer of (6 mil) polyethylene. All microbial contaminated waste should be double bagged or wrapped in (6 mil) polyethylene. The designated work area should include taping along seams, isolating HVAC system, sealing all openings, and regulated HEPA filtered intake air from outside and a HEPA negative air system exhausting the work area to outside. Provide negative air pressure within the containment area utilizing variable speed, thermal protected HEPA air filtration system with:

a. Primary two stage poly filter (35% efficiency)
b. Secondary pleated poly filter (70% efficiency)
c. Final HEPA filter (99.97% @ 0.3-micron efficiency)
d. Utilize carbon assisted HEPA air polishers for 24 hours to provide final level of microbial remediation.

e. Provide mechanical dehumidifiers of sufficient capacity to create an effective drying system as necessary.

f. Respiratory protection in accordance with the OSHA respiratory protection standard (29 CFR 1910.134), is recommended. Gloves and eye protection should be worn.

g. During cleaning, vacating people from spaces adjacent to the work area is not necessary but is recommended in the presence of infants (less than 12 months old), those recovering from recent surgery, immune suppressed people, or people with chronic inflammatory diseases (e.g., asthma, hypersensitivity pneumonitis, and severe allergies), and people diagnosed with fungal related diseases.

h. The Clinic HVAC unit and louvers should be HEPA vacuumed and cleaned using one of the recommended cleaners above, or similar, to remove settled dust and fungal spores. The vent should be isolated from active HVAC airflow during cleaning to prevent redistribution of spores to other areas of the building.

i. Inspect the area for any evidence of, condensation, or other moisture sources, and correct any deficiencies if identified.

j. After the surfaces are properly cleaned and dried, a particulate suppressant may be used to help pull any spores out of the air. After the suppressant is dried, HEPA vacuum the surfaces again.

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Please see attached laboratory results for chain of custody, complete sample location, and description information.

The samples were collected in accordance with generally accepted standards including the ACGIH and AIHA's *Field Guide for the Determination of Biological Contaminants in Environmental Samples*.

Limitations, Disclaimers, and Conditions of Use

This report is subject to the following limitations, which are a material part of its terms.

1. No Regulatory Standards Exist for Mold
No federal, state, or local agency has established numerical exposure limits for airborne mold. This is due to variability in individual immunologic sensitivity and the absence of established dose-response data. In the absence of such standards, findings in this report are interpretive only, based on the genera/species identified and comparisons between sampled environments (e.g., indoor vs. outdoor, complaint vs. non-complaint areas). Airborne fungal levels fluctuate with weather, humidity, air movement, and occupant activity; a different sampling date or condition could yield materially different results.

2. Scope of Services
This report addresses indoor air quality/mold conditions only. It does **not** address asbestos, lead-based paint, or any other environmental hazard, and no such testing was performed unless expressly stated elsewhere in this report.

3. Point-in-Time Data; No Predictive Value
Sampling results reflect conditions at the specific location(s), date, and time of sampling only. They are a snapshot and shall not be construed as representing, predicting, or warranting conditions before, after, or at any time other than the sampling event. Because inspection and sampling are inherently limited in scope, this report may not identify all conditions present at the property, including conditions in areas that were inaccessible, concealed, or not sampled (e.g., within wall cavities, sub-floors, chases, or behind finishes). No representation is made regarding inaccessible areas, and Pinnacle Consultants accepts no liability for conditions in such areas.

4. No Warranty; As-Is Basis
This report is provided on an as-is basis and is **not** a guarantee, warranty, or certification of any kind, express or implied, including without limitation any implied warranty of merchantability or fitness for a particular purpose. This report does not certify or validate the adequacy, completeness, or success of any mold remediation performed by others; information on remediation history should be obtained directly from the property owner/seller.

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5. No Medical Opinion
Nothing in this report constitutes medical advice, a diagnosis, or a determination — express or implied — as to whether any individual's exposure to conditions at the property has caused, or may in the future cause, adverse health effects. Only a licensed physician is qualified to evaluate the medical significance of any exposure. This report is not, and shall not be relied upon as, a basis for any evacuation or remediation decision; such decisions rest solely with the owner, prospective purchaser, and/or a licensed remediation contractor, in consultation with appropriate professionals.

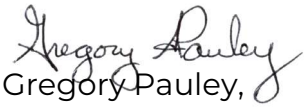
6. Intended Use and Reliance
This report is prepared exclusively for the use of the client for whom it was prepared for and Pinnacle Consultants. It may be disclosed to third parties for informational purposes only; no third party is entitled to rely on this report, and Pinnacle Consultants owes no duty of care to any party other than its client listed on the cover page. Any party choosing to rely on this report for any purpose does so entirely at its own risk. This report my be reproduced but only in full.

7. Limitation of Liability
To the fullest extent permitted by law, Pinnacle Consultants and its employees, agents, and subcontractors shall have no liability for any indirect, incidental, special, consequential, exemplary, or punitive damages — including loss of investment, value, use, or expenditure — arising from (a) use or inability to use this or any prior Pinnacle report, (b) the acts, statements, or omissions of any third party, or (c) any other matter relating to Pinnacle's services regarding the referenced property, regardless of the theory of liability (contract, tort, negligence, strict liability, or otherwise) and even if Pinnacle was advised of the possibility of such damages. **In no event shall Pinnacle Consultants' total aggregate liability to any party, arising out of or related to this report, exceed the total fees paid to Pinnacle for its preparation.**

8. Severability
If any provision of these limitations is held unenforceable, the remaining provisions shall remain in full force and effect, and the unenforceable provision shall be modified to the minimum extent necessary to make it enforceable.

Pinnacle Consultants greatly appreciates the opportunity to provide you with our mold assessment services for this project. Should you have any questions, please contact our office at your earliest convenience at 304-757-5204.

Sincerely,
Pinnacle Consultants, LLC



Gregory Pauley,
President

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LABORATORY ANALYSIS RESULTS

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HAYES MICROBIAL CONSULTING

#26048613

Analysis Report prepared for

Pinnacle Consultants, LLC

10 Independent Avenue
Nitro, WV 25143

Phone: **(304) 757-5204**

26-654
CK Elementary

Collected: **September 14, 2026**
Received: **September 15, 2026**
Reported: **September 16, 2026**

We would like to thank you for trusting Hayes Microbial for your analytical needs!
We received 18 samples by FedEx in good condition for this project on September 15th, 2026.

The results in this analysis pertain only to this job, collected on the stated date, and should not be used in the interpretation of any other job. Information supplied by the customer can affect the validity of results. These results apply only to the samples as received. This report may not be duplicated, except in full, without the written consent of Hayes Microbial Consulting, LLC.

All information provided to Hayes Microbial is confidential information relating to our customers and their clients. We will not disclose, copy, or distribute any information verbally or written, except to those designated by the customer(s). We take confidentiality very seriously. No changes to the distribution list will be made without the express consent of the customer.

This laboratory bears no responsibility for sample collection activities, analytical method limitations, or your use of the test results. Interpretation and use of test results are your responsibility. Any reference to health effects or interpretation of mold levels is strictly the opinion of Hayes Microbial. In no event, shall Hayes Microbial or any of its employees be liable for lost profits or any special, incidental or consequential damages arising out of the use of these test results.

Steve Hayes, BSMT(ASCP)
Laboratory Director
Hayes Microbial Consulting, LLC.

EPA Laboratory ID: VA01419 | Lab ID: #188863 | DPH License: #PH-0198

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26-654
CK Elementary

#26048613

Spore Trap
SOP - HMC#101

Sample Number*	1	654 - 1		2	654 - 2		3	654 - 3		4	654 - 4	
Sample Name*	238			231			224			217		
Sample Volume*	75 L			75 L			75 L			75 L		
Reporting Limit	13 spores/m³			13 spores/m³			13 spores/m³			13 spores/m³		
Background	2			2			2			2		
Fragments	ND			ND			ND			13/m³		
Organism	**Raw Count**	**Count / m³**	**% of Total**	**Raw Count**	**Count / m³**	**% of Total**	**Raw Count**	**Count / m³**	**% of Total**	**Raw Count**	**Count / m³**	**% of Total**
Alternaria												
Ascospores	1	13	100.0%				1	13	50.0%	2	27	66.7%
Aspergillus|Penicillium												
Basidiospores										1	13	33.3%
Bipolaris|Drechslera												
Chaetomium												
Cladosporium												
Curvularia				1	13	50.0%						
Epicoccum												
Fusarium												
Memnoniella												
Myxomycetes/Periconia				1	13	50.0%						
Nigrospora												
Pithomyces												
Rusts/Smuts												
Spegazzinia												
Stachybotrys												
Stemphylium												
Tetraploa												
Torula							1	13	50.0%			
Ulocladium												
Total	1	13	100%	2	26	100%	2	26	100%	3	40	100%

Water Damage Indicator | Common Allergen | Slightly Higher than Baseline | Significantly Higher than Baseline | Ratio Abnormality

* indicates data provided by the customer

Collected: **Sep 14, 2026** Received: **Sep 15, 2026** Reported: **Sep 16, 2026**

HAYES MICROBIAL CONSULTING

Project Analyst: Richard Garrett, Date: **09 - 16 - 2026**

Reviewed By: Landon Grivetti, Date: **09 - 16 - 2026**

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CK Elementary

#26048613

Spore Trap
SOP - HMC#101

Sample Number*	5	654 - 5		6	654 - 6		7	654 - 7		8	654 - 8	
Sample Name*	212			Hall @ 190			182			144		
Sample Volume*	75 L			75 L			75 L			75 L		
Reporting Limit	13 spores/m³			13 spores/m³			13 spores/m³			13 spores/m³		
Background	2			2			1			2		
Fragments	ND			ND			ND			ND		
Organism	**Raw Count**	**Count / m³**	**% of Total**	**Raw Count**	**Count / m³**	**% of Total**	**Raw Count**	**Count / m³**	**% of Total**	**Raw Count**	**Count / m³**	**% of Total**
Alternaria												
Ascospores				1	13	33.3%	1	13	100.0%	2	27	66.7%
Aspergillus|Penicillium												
Basidiospores												
Bipolaris|Drechslera												
Chaetomium												
Cladosporium												
Curvularia	1	13	100.0%									
Epicoccum												
Fusarium												
Memnoniella												
Myxomycetes/Periconia				2	27	66.7%				1	13	33.3%
Nigrospora												
Pithomyces												
Rusts/Smuts												
Spegazzinia												
Stachybotrys												
Stemphylium												
Tetraploa												
Torula												
Ulocladium												
Total	1	13	100%	3	40	100%	1	13	100%	3	40	100%

Water Damage Indicator | Common Allergen | Slightly Higher than Baseline | Significantly Higher than Baseline | Ratio Abnormality

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Spore Trap
SOP - HMC#101

Sample Number*	9	654 - 9		10	654 - 10		11	654 - 11		12	654 - 12	
Sample Name*	180			169			159			178		
Sample Volume*	75 L			75 L			75 L			75 L		
Reporting Limit	13 spores/m³			13 spores/m³			13 spores/m³			13 spores/m³		
Background	1			2			2			2		
Fragments	ND			ND			13/m³			ND		
Organism	**Raw Count**	**Count / m³**	**% of Total**	**Raw Count**	**Count / m³**	**% of Total**	**Raw Count**	**Count / m³**	**% of Total**	**Raw Count**	**Count / m³**	**% of Total**
Alternaria										1	13	12.5%
Ascospores	1	13	100.0%	2	27	66.7%	4	53	57.1%	3	40	37.5%
Aspergillus|Penicillium												
Basidiospores				1	13	33.3%	2	27	28.6%	1	13	12.5%
Bipolaris|Drechslera												
Chaetomium												
Cladosporium												
Curvularia												
Epicoccum												
Fusarium												
Memnoniella												
Myxomycetes/Periconia							1	13	14.3%	2	27	25.0%
Nigrospora												
Pithomyces												
Rusts/Smuts												
Spegazzinia												
Stachybotrys												
Stemphylium												
Tetraploa										1	13	12.5%
Torula												
Ulocladium												
Total	1	13	100%	3	40	100%	7	93	100%	8	106	100%

Water Damage Indicator | Common Allergen | Slightly Higher than Baseline | Significantly Higher than Baseline | Ratio Abnormality

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#26048613

Spore Trap
SOP - HMC#101

Sample Number*	13	654 - 13		14	654 - 14		15	654 - 15		16	654 - 16	
Sample Name*	Cafeteria / 124			111			142			Outisde		
Sample Volume*	75 L			75 L			75 L			75 L		
Reporting Limit	13 spores/m³			13 spores/m³			13 spores/m³			13 spores/m³		
Background	1			2			2			2		
Fragments	ND			ND			ND			27/m³		
Organism	**Raw Count**	**Count / m³**	**% of Total**	**Raw Count**	**Count / m³**	**% of Total**	**Raw Count**	**Count / m³**	**% of Total**	**Raw Count**	**Count / m³**	**% of Total**
Alternaria										6	80	4.4%
Ascospores	5	67	83.3%	2	27	50.0%	1	13	100.0%	62	830	45.9%
Aspergillus|Penicillium												
Basidiospores										22	290	16.3%
Bipolaris|Drechslera												
Chaetomium												
Cladosporium										36	480	26.7%
Curvularia	1	13	16.7%									
Epicoccum												
Fusarium												
Memnoniella												
Myxomycetes/Periconia				2	27	50.0%				2	27	1.5%
Nigrospora										2	27	1.5%
Pithomyces										1	13	<1%
Rusts/Smuts												
Spegazzinia										1	13	<1%
Stachybotrys												
Stemphylium												
Tetraploa												
Torula										3	40	2.2%
Ulocladium												
Total	6	80	100%	4	54	100%	1	13	100%	135	1800	100%

Water Damage Indicator | Common Allergen | Slightly Higher than Baseline | Significantly Higher than Baseline | Ratio Abnormality

* indicates data provided by the customer

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26-654
CK Elementary

#26048613

Spore Trap
SOP - HMC#101

Sample Number*	17	654 - 17	
Sample Name*	**Clinic**		
Sample Volume*	75 L		
Reporting Limit	13 spores/m³		
Background	2		
Fragments	ND		
Organism	**Raw Count**	**Count / m³**	**% of Total**
Alternaria			
Ascospores	1	13	25.0%
Aspergillus|Penicillium			
Basidiospores			
Bipolaris|Drechslera			
Chaetomium			
Cladosporium	3	40	75.0%
Curvularia			
Epicoccum			
Fusarium			
Memnoniella			
Myxomycetes/Periconia			
Nigrospora			
Pithomyces			
Rusts/Smuts			
Spegazzinia			
Stachybotrys			
Stemphylium			
Tetraploa			
Torula			
Ulocladium			
Total	4	53	100%

Water Damage Indicator | Common Allergen | Slightly Higher than Baseline | Significantly Higher than Baseline | Ratio Abnormality

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26-654
CK Elementary

#26048613

Direct Analysis
SOP - HMC#102

#18	Bio-Tape (1.00 cm2*)	Organism	Spore Estimate	Mycelial Estimate
654 - 18 - Clinic Vent Tape		Cladosporium	Moderate	Few

* indicates data provided by the customer

Collected: **Sep 14, 2026** Received: **Sep 15, 2026** Reported: **Sep 16, 2026**

HAYES MICROBIAL CONSULTING

Project Analyst: Richard Garrett, Date: **09 - 16 - 2026**

Reviewed By: Landon Grivetti, Date: **09 - 16 - 2026**

3005 East Boundary Terrace, Suite F. Midlothian, VA. 23112 (804) 562-3435 contact@hayesmicrobial.com

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Spore Trap Information

Reporting Limit	The Reporting Limit is the lowest number of spores that can be detected based on the total volume of the sample collected and the percentage of the slide that is counted. At Hayes Microbial, 100% of the slide is read so the LOD is based solely on the total volume. Raw spore counts that exceed 500 spores will be estimated.
Blanks	Results have not been corrected for field or laboratory blanks.
Background	The Background is the amount of debris that is present in the sample. This debris consists of skin cells, dirt, dust, pollen, drywall dust and other organic and non-organic matter. As the background density increases, the likelihood of spores, especially small spores such as those of Aspergillus and Penicillium may be obscured. The background is rated on a scale of 1 to 5 and each level is determined as follows: **NBD:** No background detected due to possible pump or cassette malfunction. Recollect sample. (Field Blanks will display NBD) **1** : <5% of field occluded. No spores will be uncountable. **2** : 5-25% of field occluded. **3** : 25-75% of field occluded. **4** : 75-90% of field occluded. **5** : >90% of field occluded. Suggested recollection of sample.
Fragments	Fragments are small pieces of fungal mycelium or spores. They are not identifiable as to type and when present in very large numbers, may indicate the presence of mold amplification.
Control Comparisons	There are no national standards for the numbers of fungal spores that may be present in the indoor environment. As a general rule and guideline that is widely accepted in the indoor air quality field, the numbers and types of spores that are present in the indoor environment should not exceed those that are present outdoors at any given time. There will always be some mold spores present in "normal" indoor environments. The purpose of sampling and counting spores is to help determine whether an abnormal condition exists within the indoor environment and if it does, to help pinpoint the area of contamination. Spore counts should not be used as the sole determining factor of mold contamination. There are many factors that can cause anomalies in the comparison of indoor and outdoor samples due to the dynamic nature of both of those environments.
Water Damage Indicator	**Blue:** These molds are commonly seen in conditions of prolonged water intrusion and usually indicate a problem.
Common Allergen	**Green:** Although all molds are potential allergens, these are the most common allergens that may be found indoors.
Slightly Higher than Baseline	**Orange:** The spore count is slightly higher than the outside count and may or may not indicate a source of contamination.
Significantly Higher than Baseline	**Red:** The spore count is significantly higher than the baseline count and probably indicates a source of contamination.
Ratio Abnormality	**Violet:** The types of spores found indoors should be similar to the ones that were identified in the baseline sample. Significant increases (more than 25%) in the ratio of a particular spore type may indicate the presence of abnormal levels of mold, even if the total number of spores of that type is lower in the indoor environment than it was outdoors.
Color Coding	Fungi that are present in indoor samples at levels lower than 200 per cubic meter are not color coded on the report, unless they are one of the water damage indicators.
Significant Figures	Raw counts and column totals may reflect more than 2 significant figures, but results should only be considered significant to 2 figures.

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Direct Analysis Information

Spore Estimate		Percentages
ND	None Detected	0%
Rare	Less than 10 spores	< 1%
Light	10 - 99 spores	1-10%
Moderate	100 - 999 spores	11-25%
Heavy	1000 - 9999 spores	26-50%
Very Heavy	10000 or greater spores	51-100%

Mycelial Estimate	
ND	None Detected No active growth at site.
Trace	Very small amount of Mycelium Probably no active growth at site.
Few	Some Mycelium Possible active growth at site.
Many	Large amount of Mycelium Probable active growth at site.

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Analyte Descriptions

Alternaria	**Habitat:**	Commonly found outdoors in soil and decaying plants. Indoors, it is commonly found on window sills and other horizontal surfaces.
	Health Effects:	A common allergen and has been associated with hypersensitivity pneumonitis. Alternaria is capable of producing toxic metabolites which may be associated with disease in humans or animals. Occasionally an agent of onychomycosis, ulcerated cutaneous infection and chronic sinusitis, principally in the immunocompromised patient.
Ascospores	**Habitat:**	A large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report.
	Health Effects:	Health affects are poorly studied, but many are likely to be allergenic.
Basidiospores	**Habitat:**	A common group of Fungi that includes the mushrooms and bracket fungi. They are saprophytes and plant pathogens. In wet conditions they can cause structural damage to buildings.
	Health Effects:	Common allergens and are also associated with hypersensitivity pneumonitis.
Cladosporium	**Habitat:**	One of the most common genera worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. The outdoor numbers are lower in the winter and often relatively high in the summer, especially in high humidity. The outdoor numbers often spike in the late afternoon and evening. Indoors, it can be found growing on textiles, wood, sheetrock, moist window sills and in HVAC supply ducts.
	Health Effects:	A common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.
Curvularia	**Habitat:**	They exist in soil and plant debris, and are plant pathogens.
	Health Effects:	They are allergenic and a common cause of allergic fungal sinusitis. An occasional cause of human infection, including keratitis, sinusitis, onychomycosis, mycetoma, pneumonia, endocarditis and desseminated infection, primarily in the immunocompromised.
Myxomycetes/Periconia	**Description:**	Found on decaying plant material and as a plant pathogen.
	Health Effects:	Some allergenic properties reported, but generally pose no health concerns to humans.

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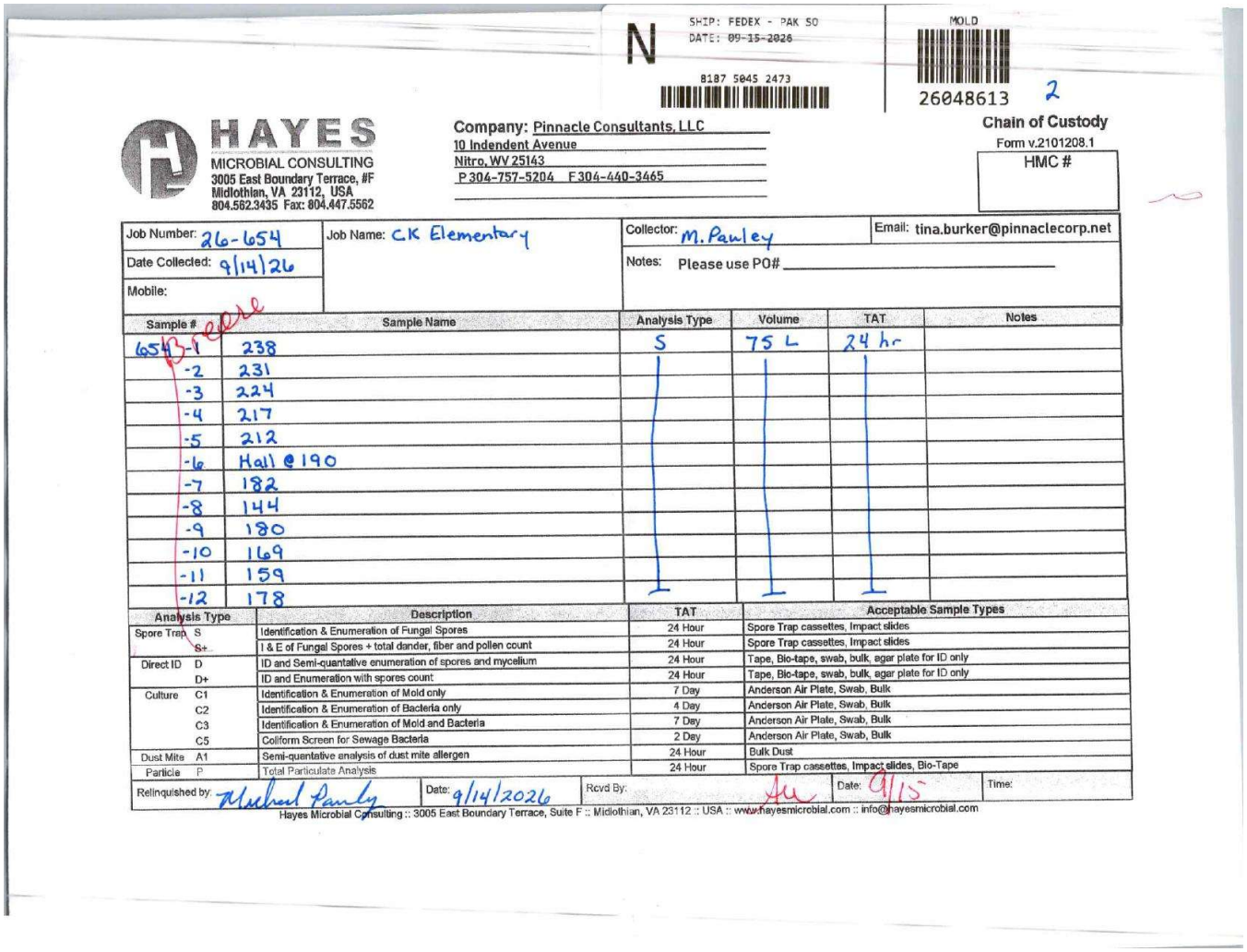
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Analyte Descriptions

Nigrospora	**Habitat:**	Found on wood, soil and decaying plant matter.
	Health Effects:	Health effects are poorly studied.
Pithomyces	**Habitat:**	Common fungus isolated from soil, decaying plant material. Rarely found indoors.
	Health Effects:	Allergenic properties are poorly studied. No cases of infection in humans.
Spegazzinia	**Habitat:**	Found in soil and occasionally on plants.
	Health Effects:	No known health effects. Allergenic properties are poorly studied.
Tetraploa	**Habitat:**	Found in soil and decaying plant material. Rarely found growing indoors.
	Health Effects:	Allergenic properties are not well studied.
Torula	**Habitat:**	Found in soil and on wood and grasses. Occasionally found growing indoors on cellulose containing materials.
	Health Effects:	A known allergen. No known cases of human infection.

CHAIN OF CUSTODY FORM

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DATE: 09-15-2026
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Chain of Custody
Form v.2101208.1
HMC #

Job Number: 26-654
Job Name: CK Elementary
Collector: M. Pauley
Email: tina.burker@pinnaclecorp.net
Date Collected: 9/14/26
Notes: Please use PO#
Mobile:

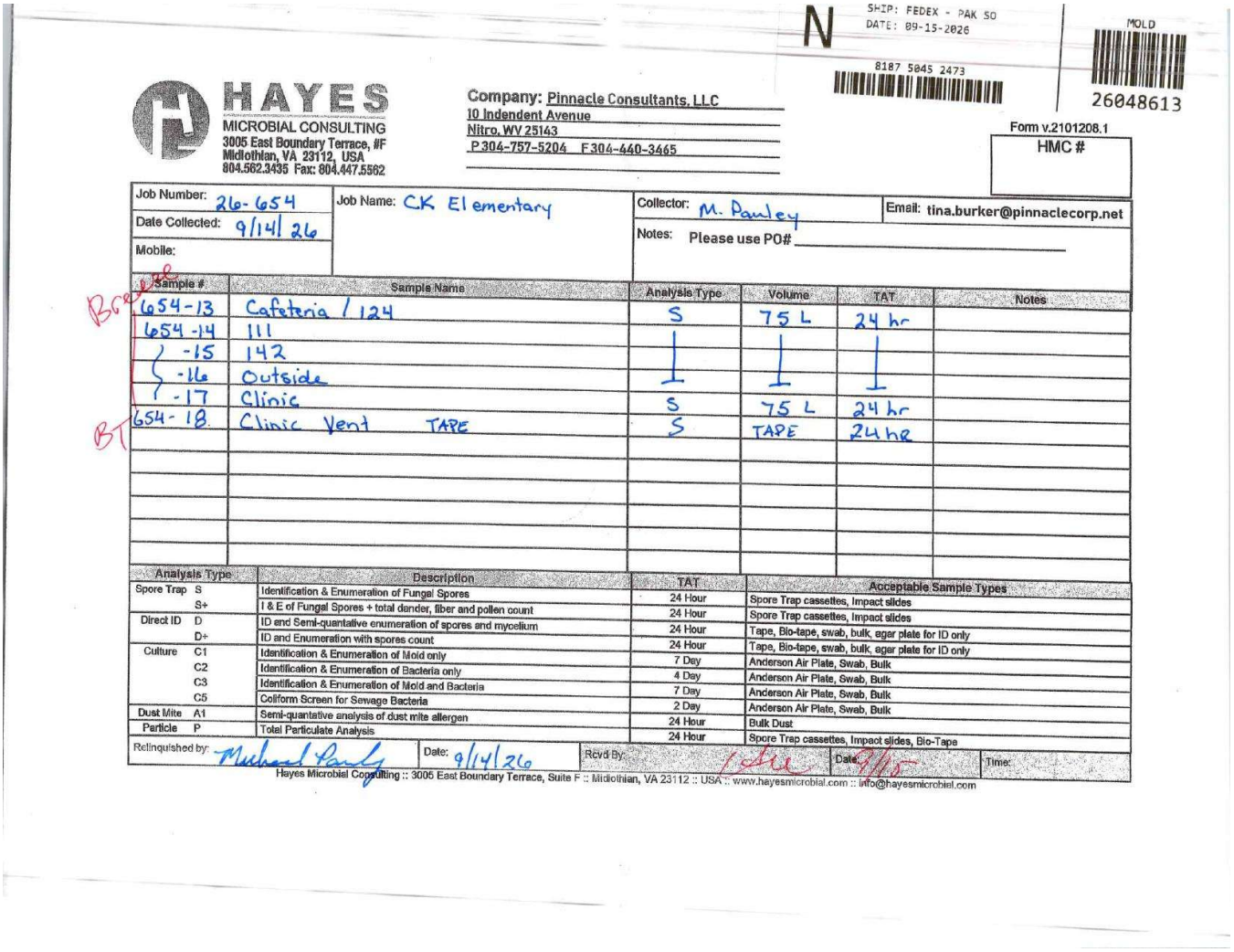
Sample #	Sample Name	Analysis Type	Volume	TAT	Notes
6543-1	238	S	75 L	24 hr	
-2	231				
-3	224				
-4	217				
-5	212				
-6	Hall @ 190				
-7	182				
-8	144				
-9	180				
-10	169				
-11	159				
-12	178				

Analysis Type		Description	TAT	Acceptable Sample Types
Spore Trap	S	Identification & Enumeration of Fungal Spores	24 Hour	Spore Trap cassettes, Impact slides
	S+	I & E of Fungal Spores + total dander, fiber and pollen count	24 Hour	Spore Trap cassettes, Impact slides
Direct ID	D	ID and Semi-quantative enumeration of spores and mycelium	24 Hour	Tape, Bio-tape, swab, bulk, agar plate for ID only
	D+	ID and Enumeration with spores count	24 Hour	Tape, Bio-tape, swab, bulk, agar plate for ID only
Culture	C1	Identification & Enumeration of Mold only	7 Day	Anderson Air Plate, Swab, Bulk
	C2	Identification & Enumeration of Bacteria only	4 Day	Anderson Air Plate, Swab, Bulk
	C3	Identification & Enumeration of Mold and Bacteria	7 Day	Anderson Air Plate, Swab, Bulk
	C5	Coliform Screen for Sewage Bacteria	2 Day	Anderson Air Plate, Swab, Bulk
Dust Mite	A1	Semi-quantative analysis of dust mite allergen	24 Hour	Bulk Dust
Particle	P	Total Particulate Analysis	24 Hour	Spore Trap cassettes, Impact slides, Bio-Tape

Relinquished by: Michael Pauley Date: 9/14/2026
Rcvd By: Date: 9/15 Time:

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DATE: 09-15-2026

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Form v.2101208.1
HMC #

Job Number: 26-654
Job Name: CK Elementary
Collector: M. Pauley
Email: tina.burker@pinnaclecorp.net
Date Collected: 9/14/26
Mobile:
Notes: Please use PO#

Sample #	Sample Name	Analysis Type	Volume	TAT	Notes
654-13	Cafeteria / 124	S	75 L	24 hr	
654-14	111	"	"	"	
-15	142	"	"	"	
-16	Outside	"	"	"	
-17	Clinic	S	75 L	24 hr	
654-18	Clinic Vent TAPE	S	TAPE	24 hr	

Analysis Type		Description	TAT	Acceptable Sample Types
Spore Trap	S	Identification & Enumeration of Fungal Spores	24 Hour	Spore Trap cassettes, Impact slides
	S+	I & E of Fungal Spores + total dander, fiber and pollen count	24 Hour	Spore Trap cassettes, Impact slides
Direct ID	D	ID and Semi-quantative enumeration of spores and mycelium	24 Hour	Tape, Bio-tape, swab, bulk, ager plate for ID only
	D+	ID and Enumeration with spores count	24 Hour	Tape, Bio-tape, swab, bulk, ager plate for ID only
Culture	C1	Identification & Enumeration of Mold only	7 Day	Anderson Air Plate, Swab, Bulk
	C2	Identification & Enumeration of Bacteria only	4 Day	Anderson Air Plate, Swab, Bulk
	C3	Identification & Enumeration of Mold and Bacteria	7 Day	Anderson Air Plate, Swab, Bulk
	C5	Coliform Screen for Sewage Bacteria	2 Day	Anderson Air Plate, Swab, Bulk
Dust Mite	A1	Semi-quantative analysis of dust mite allergen	24 Hour	Bulk Dust
Particle	P	Total Particulate Analysis	24 Hour	Spore Trap cassettes, Impact slides, Bio-Tape

Relinquished by: Michael Pauley Date: 9/14/26 Rcvd By: Date: 9/15 Time:

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CLIENT:
INSPECTOR: MP / GP
LOCATION: CK Elementary
DATE: 9/14/26
PROJECT NUMBER: 26-654

Turn-around Time:

IAQ READINGS

Client ID	Room # or Description	Temperature (°F)	Relative Humidity (%)	Carbon Dioxide (CO^2)	Sample Volume
654-1	Rm 238	71.6	52.2	814	~~75~~ 752
-2	Rm 231 - Janitor Sweeping	70.7	57.8	736	
-3	Rm 224	70.9	55.2	737	
-4	Rm 217	70.7	46.1	542	
-5	Rm 212	71.2	48.5	648	
-6	~~190~~ in Hall @ 190	71.1	51	844	
-7	182	71.7	57.1	1566	
-8	144	~~70.8~~ 72.4	~~56.0~~ 46.9	~~1606~~ 741	
-9	~~179~~ 180	70.9	57.8	1478	
-10	169	70.8	56	1606	
-11	159	~~73.1~~ 72.3	~~46.9~~ 58.3	~~741~~ 1755	
-12	178 - Library Containment	76.8	52.6	990	
-13	Cafeteria / 124	67.1	50.1	624	

Relinquished by:_____

Received by:_____

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PINNACLE CONSULTANTS

CLIENT: _____
INSPECTOR: _____
LOCATION: _____
DATE: _____
PROJECT NUMBER: _____

Turn-around Time:

IAQ READINGS

Client ID	Room # or Description	Temperature (°F)	Relative Humidity (%)	Carbon Dioxide (CO^2)	Sample Volume
-14	111	67.5	58.9	538	
-15	142	73.5	47.5	965	
-16		87.7	38.7	466	

Relinquished by: _____

Received by: _____

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FUNGAL LEVELS and FUNGAL DESCRIPTIONS

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Fungal Levels

The table below summarizes the International Institute for Building Biology & Ecology (IIBE) Healthy Home Standard thresholds used to compare indoor spore-trap results against the exterior comparison sample collected at this property. An indoor result is classified as "Unlikely," "Possible," or "Likely" to represent an indoor fungal reservoir for each metric, based on how the indoor value compares to the outdoor value.

Metric	Unlikely if Indoor Air Is:	Possible if Indoor Air Is:	Likely if Indoor Air Is:
Outdoor-Type Spores	< 1.2x Total Outdoor Air	< 2x Total Outdoor Air	> 2x Total Outdoor Air
Aspergillus/Penicillium-like	< Outdoor Air + 300	< Outdoor Air + 800	> Outdoor Air + 800
Stachybotrys	< Outdoor Air	< Outdoor Air + 10	> Outdoor Air + 10
Chaetomium	< Outdoor Air	< Outdoor Air + 20	> Outdoor Air + 20
Mycelium Fragments	< Outdoor Air + 150	< Outdoor Air + 300	> Outdoor Air + 300
Sum of Diverse Spore Types	< Outdoor Air + 400	< Outdoor Air + 800	> Outdoor Air + 800

Source: International Institute for Building Biology & Ecology (IBE) Healthy Home Standard, Version 1, Copyright 2010.

Based on this standard, none of the indoor air samples collected at this property met or exceeded the "Possible" or "Likely" thresholds for any metric when compared to the exterior sample. However, the tape-lift (direct exam) sample collected from the Clinic supply vent is not diluted by this air-sample comparison and is discussed separately below; it identified a Moderate spore estimate of *Cladosporium* with a Few (possible active growth) mycelial estimate.

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Fungal Descriptions

The following describes the fungal genera and spore types identified in the air and tape-lift samples collected at this property.

Alternaria: Commonly found outdoors in soil and decaying plant material, and often found indoors on window sills and other horizontal surfaces. It is a common allergen and has been associated with hypersensitivity pneumonitis in sensitive individuals.

Ascospores: A large and diverse group representing more than 3,000 species of fungi produced by sac fungi (Ascomycetes). Outdoor counts often rise sharply following rain, and elevated counts are typically reflective of outdoor conditions rather than an indoor source. Health effects are poorly studied, though many species are considered allergenic.

Basidiospores: Spores produced by mushrooms, bracket fungi, and related organisms (Basidiomycetes). These are common allergens associated with decaying wood and outdoor vegetation, and in prolonged wet conditions can contribute to structural damage.

Cladosporium: One of the most common mold genera worldwide, found in soil, plant debris, and on the leaf surfaces of living plants outdoors. Indoors, it can grow on textiles, wood, drywall, moist window sills, and HVAC components, particularly where dust and condensation accumulate. It is a well-recognized allergen, producing more than ten allergenic antigens and a common cause of hypersensitivity pneumonitis.

Curvularia: Found in soil and plant debris outdoors and considered a plant pathogen. It is allergenic and a recognized cause of allergic fungal sinusitis, with occasional infections reported primarily in immunocompromised individuals.

Myxomycetes/Periconia: A grouping of slime molds (Myxomycetes) and a dematiaceous mold (Periconia) that share similar spore morphology under microscopic analysis. These are primarily outdoor organisms found on decaying plant material, and while some allergenic properties have been reported, they generally pose little health concern to humans.

Nigrospora: Found on wood, soil, and decaying plant matter outdoors. Health effects are poorly studied, and it is considered to have little indoor significance unless found in unusually high concentrations relative to outdoor air.

Pithomyces: A common fungus isolated from soil and decaying plant material outdoors; it is rarely found growing indoors. Allergenic properties are poorly studied, and no cases of human infection have been documented.

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Spegazzinia: Found in soil and occasionally on plant surfaces outdoors. No known health effects have been documented, and its allergenic properties are poorly studied.

Tetraploa: Found in soil and decaying plant material outdoors; it is rarely found growing indoors. Its allergenic properties are not well studied.

Torula: A dematiaceous mold found in soil and on wood and grasses outdoors, occasionally identified indoors growing on cellulose-containing materials. It is a known allergen, though no cases of human infection have been documented.